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ISOGINSENOSIDE-Rh₃, A NEW TRITERPENOID SAPONIN FROM THE FRUITS OF *PANAX GINSENG* C. A. MEY

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A new dammarane-type triterpene monoglucoside, named isoginsenoside-Rh₃, has been isolated from the fruits of *Panax ginseng* C. A. Mey, together with eight known analogs, ginsenoside-Rb₁, -Rb₂, -Rc, -Rd, -Re, -Rg₁, -Rh₁, -Rh₂. On the basis of chemical and physicochemical evidence, the structure of isoginsenoside-Rh₃ has been elucidated as 3-*O*-β-D-glucopyranosyl-dammarane-(*E*)-20(22),24-diene-3β,12β-diol (**1**).

Keywords: *Panax ginseng* C. A. Mey; Fruits; Triterpenoid; Isoginsenoside-Rh₃

INTRODUCTION

There is growing evidence in the literature that the fruits of *Panax ginseng* C. A. Mey, the well-known traditional herbal remedy used in Chinese medicine, possess an array of interesting pharmacological actions, such as cardioprotection, vasorelaxant, antistress, a stimulating activity of the central nervous system with effects on memory, learning and behaviour. The biologically active constituents of ginseng fruits have been studied extensively and various ginsenosides, dammarane-type triterpene oligoglycosides, have been characterized as the principal ingredients [1–3]. As a part of elucidating the biologically active principles of the fruits of *P. ginseng*, we describe the structural determination of a new dammarane-type triterpene monoglucoside named as isoginsenoside-Rh₃ (**1**).

RESULTS AND DISCUSSION

1 was isolated as white powder. It gave positive Liebermann–Burchard and Molish reactions, and electro-spray ionization mass spectrometry (ESI-MS) showed quasi-molecular ion peaks

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at m/z 627 $[M + Na]^+$ and 443 $[M - \text{glucosyl} + H]^+$. Its molecular formula $C_{36}H_{60}O_7$ was determined by high-resolution secondary-ion mass spectrometry (HR-SIMS), m/z 627.4225 $[M + Na]^+$. The IR spectrum displayed strong absorption bands at 3407 and 1075 cm^{-1} , suggestive of the glycosidic structure. Characteristic signals owing to a glucosyl group were observed in the NMR spectra of **1** [^1H NMR δ (ppm): 4.01 (1H, dd, $J = 6.0, 8.5$ Hz, glc-H-5), 4.03 (1H, t, $J = 8.5$ Hz, glc-H-2), 4.21 (1H, t, $J = 8.5$ Hz, glc-H-4), 4.25 (1H, t, $J = 8.5$ Hz, glc-H-3), 4.40 (1H, dd, $J = 6.0, 11.5$ Hz, glc-Ha-6), 4.59 (1H, d, $J = 11.5$ Hz, glc-Hb-6), 4.95 (1H, d, $J = 8.0$ Hz, glc-H-1); ^{13}C NMR δ (ppm): 106.9 (C-1), 75.8 (C-2), 78.7 (C-3), 71.8 (C-4), 78.4 (C-5), 63.1 (C-6)]. Acid hydrolysis of **1** with 9% methanolic HCl yielded an aglycone and one sugar which were identified as D-glucose by PC comparison with authentic sample. The large J (8.0 Hz) indicated β -glucosidic linkages.

Eight methyl groups [^1H NMR δ (ppm): 0.79 (s), 0.96 (s), 0.98 (s), 1.00 (s), 1.31 (s), 1.56 (s), 1.60 (s), 1.80 (s); ^{13}C NMR δ : 13.1, 15.8, 16.5, 16.8, 17.0, 17.7, 25.7, 28.1] and two olefinic bonds signals [^1H NMR δ : 5.49 (1H, t, $J = 7.0$ Hz, H-22), 5.21 (1H, t, $J = 6.8$ Hz, H-24) ^{13}C NMR δ : 123.8 d, 124.5 d, 131.2 s, 140.1 s] were observed. All these data suggested that **1** is a dammarane-type triterpenoid glycoside with double bonds [4,5]. Compared with ginsenoside Rg₆ (**2**) [4] and ginsenoside Rh₃ (**3**) [5], **1** was similar to **2**, except for the signals assigned to cycles A, B, C, D and sugar moieties (Fig. 1), and similar to **3** except for

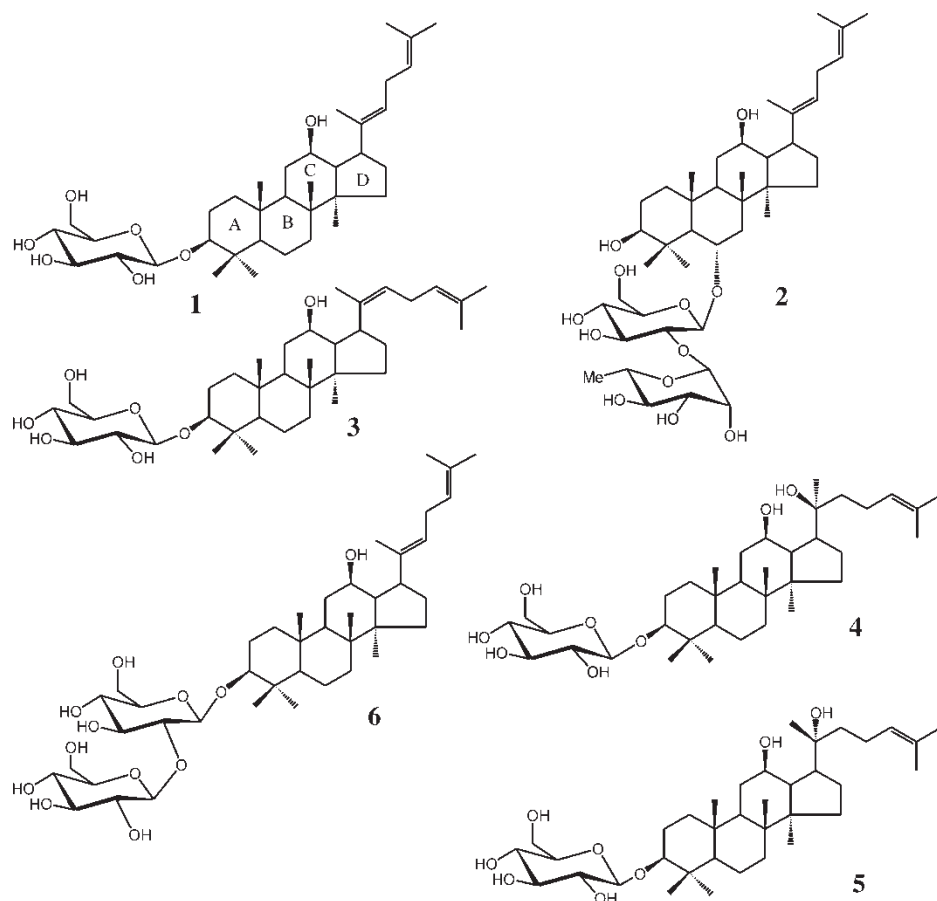


FIGURE 1 Structures of compounds 1–6.

the signals assigned to the C₁₇ side-chain in the ¹³C NMR spectral data. These results indicated that **1** is closely related to **2** and **3**. Further comparison of the ¹H and ¹³C NMR data of **1** with those of **2** [4], **3** [5], 20(*S*)-ginsenoside Rh₂ (**4**) [6], 20(*R*)-ginsenoside Rh₂ (**5**) [5], and ginsenoside-Rg₅ (**6**) [7] (Fig. 1, and Table I) showed that the signals of the C₁₇ side-chain are similar to those of **2** and **6**. The stereochemistry of the double bond at C-20(22) was proposed to be *E* from the C₂₁-methyl signal at δ 13.0 [4,7] in the ¹³C NMR spectrum. These results suggest that **1** is 3-*O*-β-D-glucopyranosyl-dammarane-(*E*)-20(22),24-diene-3β,12β-diol (Fig. 1), a novel compound, named isoginsenoside-Rh₃.

Meanwhile, the ¹H and ¹³C NMR signals of **1** were assigned by DEPT and 2D NMR with pulse-field gradient (PFG) techniques (gCOSY, gHMQC and gHMBC).

TABLE I ¹³C NMR spectral data (δ in C₅D₅N) of **1**–**6**

| Carbon no. | Compounds* | | | | | |
|------------|------------|--------------|--------------|--------------|--------------|--------------|
| | 1 | 2 [4] | 3 [5] | 4 [6] | 5 [5] | 6 [7] |
| 1 | 39.2 t | 39.6 t | 39.3 | 39.4 | 39.1 | 39.2 |
| 2 | 26.7 t | 27.6 t | 27.0 | 27.3 | 26.7 | 28.0 |
| 3 | 88.7 d | 78.3 d | 88.3 | 88.9 | 88.8 | 88.8 |
| 4 | 39.7 s | 39.6 s | 40.3 | 40.3 | 40.1 | 40.1 |
| 5 | 56.4 d | 60.7 d | 56.4 | 56.7 | 56.4 | 56.3 |
| 6 | 18.4 t | 74.1 d | 18.5 | 18.7 | 18.5 | 18.3 |
| 7 | 35.3 t | 46.1 t | 35.3 | 36.0 | 35.2 | 35.2 |
| 8 | 40.2 s | 41.3 s | 37.1 | 37.2 | 37.0 | 39.6 |
| 9 | 50.7 d | 50.2 d | 50.9 | 50.7 | 50.4 | 50.7 |
| 10 | 37.0 s | 39.4 s | 39.7 | 39.8 | 39.7 | 36.9 |
| 11 | 32.2 t | 32.1 t | 32.2 | 32.2 | 32.2 | 32.1 |
| 12 | 72.4 d | 72.5 d | 71.9 | 71.1 | 70.9 | 72.5 |
| 13 | 50.8 d | 50.6 d | 50.4 | 48.8 | 49.2 | 50.3 |
| 14 | 50.9 s | 50.8 s | 51.2 | 51.9 | 51.8 | 50.9 |
| 15 | 32.6 t | 32.5 t | 32.6 | 31.5 | 31.4 | 32.5 |
| 16 | 26.7 t | 27.6 t | 26.8 | 26.8 | 26.7 | 26.6 |
| 17 | 50.4 d | 50.0 d | 51.2 | 54.8 | 50.6 | 50.8 |
| 18 | 15.8 q | 17.6 q | 16.8 | 16.8 | 16.8 | 16.4 |
| 19 | 16.8 q | 17.6 q | 16.5 | 16.4 | 16.4 | 16.5 |
| 20 | 140.1 s | 140.0 s | 140.2 | 73.2 | 73.0 | 140.1 |
| 21 | 13.1 q | 13.0 q | 27.4 | 27.0 | 22.8 | 13.1 |
| 22 | 123.8 d | 123.0 d | 123.8 | 35.4 | 43.3 | 123.2 |
| 23 | 27.4 t | 27.4 t | 30.0 | 23.1 | 22.6 | 27.4 |
| 24 | 124.5 d | 123.7 d | 125.4 | 126.4 | 126.1 | 123.5 |
| 25 | 131.2 s | 131.2 s | 131.5 | 130.7 | 130.8 | 131.2 |
| 26 | 25.7 q | 25.6 q | 25.7 | 25.7 | 25.8 | 25.6 |
| 27 | 17.7 q | 16.8 q | 17.7 | 17.7 | 17.7 | 17.7 |
| 28 | 28.1 q | 32.1 q | 28.2 | 28.3 | 28.2 | 28.7 |
| 29 | 16.5 q | 17.5 q | 15.8 | 16.0 | 15.8 | 15.7 |
| 30 | 17.0 q | 17.1 q | 17.0 | 17.3 | 17.4 | 16.9 |
| 1' | 106.9 d | 101.7 d | 106.9 | 106.7 | 106.9 | 105.0 |
| 2' | 75.8 d | 79.3 d | 75.7 | 75.8 | 75.8 | 83.3 |
| 3' | 78.7 d | 78.4 d | 78.7 | 78.7 | 78.8 | 78.1 |
| 4' | 71.8 d | 72.5 d | 71.9 | 72.2 | 71.9 | 71.5 |
| 5' | 78.4 d | 78.4 d | 78.3 | 78.0 | 78.3 | 77.8 |
| 6' | 63.1 t | 63.0 t | 63.1 | 63.3 | 63.1 | 62.6 |
| 1'' | | 101.8 d | | | | 105.9 |
| 2'' | | 72.2 d | | | | 77.0 |
| 3'' | | 72.3 d | | | | 78.2 |
| 4'' | | 74.3 d | | | | 71.5 |
| 5'' | | 69.4 d | | | | 78.0 |
| 6'' | | 18.7 q | | | | 62.7 |

* **1**, isoginsenoside Rh₃; **2**, ginsenoside Rg₆; **3**, ginsenoside Rh₃; **4**, 20(*S*)-ginsenoside Rh₂; **5**, 20(*R*)-ginsenoside Rh₂; **6**, ginsenoside Rg₅.

EXPERIMENTAL

General Experimental Procedures

IR spectra were recorded on a Thermo Nicolet Nexus 470 FT-IR Spectrometer with KBr pellets. Optical rotations were determined on a Perkin-Elmer 243 Polarimeter. NMR spectra were obtained on a Varian INOVA-500 spectrometer in pyridine- d_5 at 500 MHz for ^1H and 125 MHz for ^{13}C . Chemical shifts (δ ppm) are given relative to TMS as internal standard. ESI-TOF-MS and HR-SIMS spectra were recorded on MDS SCIEX API QSTAR and APEX II FT-ICR (Bruker Daltonics) mass spectrometers, respectively. Macroporous resin Diaion-101 was produced by Nankai University of China.

Plant Material

The fresh fruits of *Panax ginseng* C. A. Mey. were collected in Changbai Country of Jilin Province, and were identified by Profesor Xiang-gao Li. The sarcocarp was taken from the fresh fruits and stored at 0°C before extraction.

Extraction and Isolation

The fresh fruits (10 kg) of *Panax ginseng* were mixed with water (50 L) to yield a syrup solution and then the seeds were removed. The syrup solution was centrifuged to get a supernatant. After evaporation of water solution *in vacuo*, the residue (480 g) was chromatographed over the Diaion-101 resin, eluting with H_2O (30 L) and 70% EtOH (30 L), respectively. Removal of the EtOH from the 70% EtOH fraction under reduced pressure gave a water solution that was extracted with n-BuOH saturated with H_2O to afford n-BuOH extracts (80 g). The latter extracts were further purified by silica gel column chromatography using CHCl_3 -MeOH (10 : 2 \rightarrow 10 : 2.5 \rightarrow 10 : 3 \rightarrow 10 : 4 \rightarrow 10 : 5) as solvent and then repeatedly chromatographed on silica gel column under low pressure to yield isoginsenoside-Rh₃ (**1**) (31 mg), ginsenosides-Rh₂ (10 mg), -Rh₁ (12 mg), -Rg₁ (40 mg), -Re (45 mg), -Rd (11 mg), -Rc (8 mg), -Rb₂ (16 mg), -Rb₁ (32 mg), respectively.

Isoginsenoside-Rh₃ (**1**)

A white powder that gave positive Liebermann-Burchard and Molish reactions; $[\alpha]_{\text{D}} + 25.6$ (c, 1.0, MeOH); IR ν_{max} (KBr) (cm^{-1}): 3407 (OH), 1636 (C=C), 1453, 1385 ($-\text{CH}_3$), 1075 (O-gly), 1029; ESI-TOF-MS: m/z 627 $[\text{M} + \text{Na}]^+$, 443 $[\text{M} - \text{glucosyl} + \text{H}]^+$; HR-SIMS: m/z 627.4225 $[\text{M} + \text{Na}]^+$ (calcd for $\text{C}_{36}\text{H}_{60}\text{O}_7\text{Na}$ 627.4231); ^1H NMR (500 MHz, py- d_5) δ (ppm): 0.74 (1H, d, $J = 12.0$ Hz, H-5), 0.79 (3H, s, Me-19), 0.96 (3H, s, Me-30), 0.98 (3H, s, Me-29), 1.00 (3H, s, Me-18), 1.31 (3H, s, Me-28), 1.56 (3H, s, Me-27), 1.60 (3H, s, Me-26), 1.80 (3H, s, Me-21), 2.77 (2H, dd, $J = 7.0, 12.0$ Hz, H-23), 2.81 (1H, m, H-17), 3.37 (1H, d.t., $J = 3.5, 8.0$ Hz, H-3), 3.90 (1H, m, H-12), 4.01 (1H, dd, $J = 6.0, 8.5$ Hz, glc-H-5), 4.03 (1H, t, $J = 8.5$ Hz, glc-H-2), 4.21 (1H, t, $J = 8.5$ Hz, glc-H-4), 4.25 (1H, t, $J = 8.5$ Hz, glc-H-3), 4.40 (1H, dd, $J = 6.0, 11.5$ Hz, glc-Ha-6), 4.59 (1H, d, $J = 11.5$ Hz, glc-Hb-6), 4.95 (1H, d, $J = 8.0$ Hz, glc-H-1), 5.21 (1H, t, $J = 7.0$ Hz, H-24), 5.49 (1H, t, $J = 7.0$ Hz, H-22); ^{13}C NMR data see in Table I.

Acid Hydrolysis of **1**

Compound **1** (10 mg) was dissolved in MeOH (10 ml) and refluxed with 9% HCl (3 ml) on a water bath for 5 h. The reaction mixture was concentrated and then dissolved in MeOH (1 ml)

for PC (n-BuOH–EtOH–H₂O–conc. NH₄OH, 45:5:49:1) together with authentic D-glucose ($R_f = 0.19$).

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